

# The Role of Aneuploidy in Cancer Evolution

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Chromosomal aberrations during cell division represent one of the first recognized features of human cancer cells, and modern detection methods have revealed the pervasiveness of aneuploidy in cancer. The ongoing karyotypic changes brought about by chromosomal instability (CIN) contribute to tumor heterogeneity, drug resistance, and treatment failure. Whole-chromosome and segmental aneuploidies resulting from CIN have been proposed to allow “macroevolutionary” leaps that may contribute to profound phenotypic change. In this review, we will outline evidence indicating that aneuploidy and CIN contribute to cancer evolution.

Intratumor heterogeneity is increasingly recognized as a ubiquitous feature across human tumors. Contributing to this heterogeneity are an increased mutation rate because of various defects in DNA damage detection and repair (genomic instability or GIN), along with gross chromosomal rearrangements and deviations from the euploid chromosome number (aneuploidy). Cancer cells derived from solid tumors almost invariably display a high rate of chromosome segregation errors, called chromosomal instability (CIN), which leads to aneuploidy. This feature was recognized early on in human tumors (see Boveri 2008, for an English translation), but its importance has been obscured by the complexity of the cancer phenotype. Multiple mechanisms have been described to account for the high rate of whole-chromosome segregation errors (wCIN) and structural aberrations (sCIN) in cancer and have been comprehensively

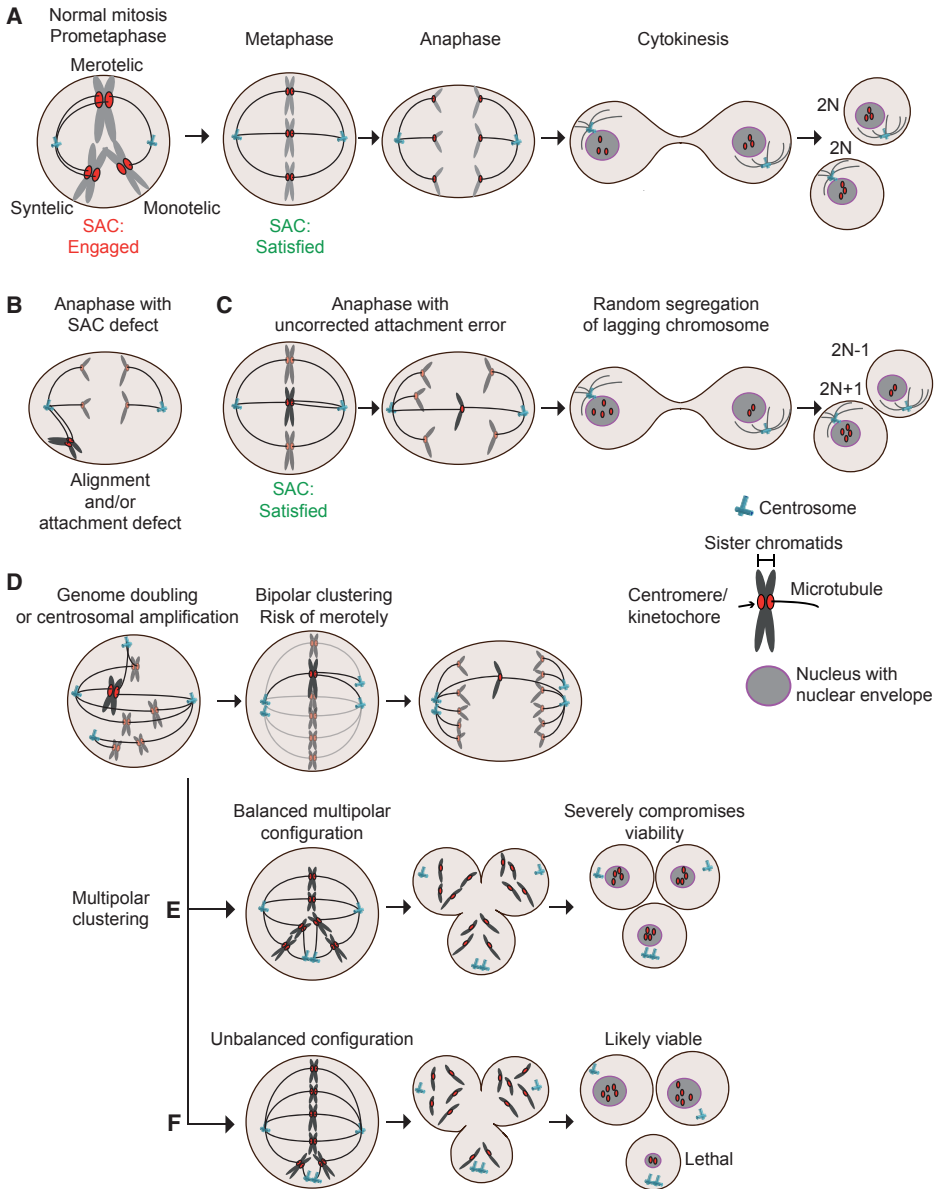
discussed elsewhere (Lengauer et al. 1998; Ganem et al. 2007; Holland and Cleveland 2009; Burrell et al. 2013b; Duijf and Benezra 2013). Here, we will briefly summarize some of the key defects leading to CIN, emphasize the importance of phenotypes allowing the expansion of highly aneuploid cells, and discuss its impact on tumor progression and treatment.

## CAUSES OF ANEUPLOIDY

### Mitotic Checkpoint Defects

The spindle assembly checkpoint (SAC) or mitotic checkpoint monitors kinetochore attachment to the mitotic spindle and delays sister chromatid separation until biorientation is achieved at metaphase (Fig. 1) (Musacchio and Salmon 2007). Heterozygous mice for SAC genes, such as BUB1, BUBR1 (BUB1B),

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**Figure 1.** Mitotic defects leading to aneuploidy. (A) Kinetochore–microtubule attachment configurations and stages of mitosis. In early mitosis, improper attachments (syntelic and monotelic) activate the spindle assembly checkpoint (SAC) to delay anaphase onset. Merotelic misattachments poorly activate the checkpoint, but error correction mechanisms contribute to the formation of correct (amphitelic) bipolar attachments prior to chromosome separation at anaphase. (B) Cells with compromised SAC activity can enter anaphase with various attachment defects, here shown with syntelic chromosomes that did not congress to the metaphase plate. (C) Uncorrected merotelic attachments lead to lagging chromosomes at anaphase onset, causing random segregation, which can result in aneuploid daughter cells. (D) Cells with centrosome amplification and tetraploid cells enter mitosis with more than two centrosomes, creating a transient multipolar spindle. Bipolar clustering creates merotelic attachments because of the random clustering of the extra centrosomes. (E) Extra centrosome can result in a multipolar anaphase creating daughter cells lacking several chromosomes in a random fashion. (F) Random clustering can create a functional spindle pole that attaches only to few chromosomes, potentially resulting in viable aneuploidy in two out of three cells. Examples in D and E are especially well tolerated by tetraploid cells.

BUB3, MAD1, MAD2, and CENPE, all display aneuploidy to various degrees (reviewed in Giam and Rancati 2015; Simon et al. 2015). SAC impairment has been linked to increased aneuploidy and in many mouse models associated with increased tumor development (either spontaneous, following drug-induced tumorigenesis or when crossed into a sensitized background). Biallelic mutations in BUB1B are linked to mosaic variegated aneuploidy (MVA) syndrome (Hanks et al. 2004). Nevertheless, it is under debate whether SAC defects represent a common CIN driver because SAC gene mutations are rarely found in cancer (discussed in the section Chromosome Cohesion Defects). In fact, some SAC proteins are frequently overexpressed in cancer, such as Mad2 (MAD2L1), which is an E2F transcriptional target aberrantly expressed in Rb mutant cells (Hernando et al. 2004). MAD2 transgenic mice develop aneuploid tumors and it was therefore proposed that SAC hyperactivation was causing CIN in this model (Sotillo et al. 2007; Kato et al. 2011). However, tetraploidization and hyperstabilization of kinetochore–microtubule (k-MT) attachments are observed upon Mad2 overexpression, which could explain how it drives CIN (discussed in the next two sections) (Hernando et al. 2004; Sotillo et al. 2007; Kabeche and Compton 2012). Another SAC protein, namely, Mad1 (MAD1L1), is overexpressed in cancer but in this case produces a dominant-negative effect and impairs SAC signaling (Ryan et al. 2012). Many SAC proteins also have roles outside mitosis that may also contribute to tumorigenesis.

### Microtubule Attachment Defects

In mitosis, spindle microtubules capture chromosomes at the kinetochore, a complex protein structure built around the centromeric region. Correct k-MT attachment is crucial for faithful chromosome segregation, and kinetochores from sister chromatids must be attached to opposite poles (Fig. 1). Aurora kinase B (AURKB) activity at the inner centromere (along with the microtubule depolymerizing protein KIF2B and KIF2C [MCAK]) plays a central role in the establishment of correct (amphitelic) at-

tachments (Fig. 1A,B). This pathway functions from prometaphase to metaphase and destabilize syntelic attachments (in which both sister kinetochores are attached to the same pole) and merotelic attachments (in which one of the kinetochores is attached to both pole) (Fig. 1A). Merotelic attachments in particular are thought to constitute a major cause of CIN because this configuration satisfies the requirements of the SAC (MT attachment and interkinetochore tension), resulting in lagging chromosomes at anaphase (Fig. 1C). Cyclin A was also shown to destabilize k-MT and promote error correction during mitotic progression, giving rise to a model whereby cyclin A degradation as cells advance from prometaphase to metaphase gradually stabilizes k-MT attachments (Kabeche and Compton 2013; Godek et al. 2015). The rate of cyclin A degradation during prometaphase might thus establish a window during which erroneous attachments can be corrected, and variations in mitotic duration could potentially compromise error correction. Impaired k-MT dynamics (stabilization) appears to contribute to CIN following deregulation of several mitotic proteins, and was also proposed to underpin CIN in adenomatous polyposis coli (APC) mutant cells (reviewed in Godek et al. 2015). Enhanced microtubule assembly rates is also a feature shared by CIN colorectal cancer cell lines and also results in microtubule hyperstabilization and chromosome segregation errors (Ertych et al. 2014). Reducing microtubule polymerization rates using a low dose of taxol or by RNAi against the microtubule stabilizer Ch-TOG reduced classical features of CIN such as lagging chromosomes and karyotypic variation. Proper activity of the CHK2-BRCA1 pathways seems important to limit AURKA activity in controlling MT assembly rates (Ertych et al. 2014, 2016).

### Centrosome and Mitotic Spindle Defects

Centrosome amplification is a widespread feature of human tumors associated with CIN and is thought to contribute to tumorigenesis through several mechanisms (Pihan et al. 1998; Ghadimi et al. 2000; Nigg and Stearns 2011; Godinho et al. 2014). Several defects result in

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supernumerary centrosomes, either directly through centriole overduplication (Nigg and Stearns 2011) or indirectly following an aborted mitosis resulting in genome doubling (tetraploidization). Extra centrosomes randomly attach to chromosomes early in mitosis, creating a transient multipolar spindle, and tend to cluster into two poles before division (Fig. 1D). Random clustering consequently increases the probability of creating merotelic attachments, and lagging chromosomes are frequent in cells with supernumerary centrosomes (Kwon et al. 2008; Ganem et al. 2009). Tetraploid cells resulting from cytokinesis failure display CIN and spontaneously generate tumors when injected in mice, whereas tetraploid cells that have lost their extra centrosomes are karyotypically stable (Fujiwara et al. 2005; Ganem et al. 2009). Genome-doubling events are also common in cancer and appear to be a precursor of CIN (Zack et al. 2013; Dewhurst et al. 2014).

Failure to cluster extra centrosomes into two poles results in multipolar divisions, likely to generate severe karyotypic changes in daughter cells. Balanced multipolar divisions are therefore lethal and very unlikely to efficiently propagate CIN, whereas an unbalanced multipolar division cell could result in the missegregation or loss of few chromosomes (Fig. 1E,F) (Ganem et al. 2007, 2009; Kwon et al. 2008; Gisselsson et al. 2010). Knockdown of the kinesin KIFC1 (HSET) by siRNA prevents clustering and results in multipolar divisions and lethality specifically in cells with multiple centrosomes. HSET small molecule inhibitors therefore represent an attractive avenue to target cancer cells (Kwon et al. 2008; Raab et al. 2012; Watts et al. 2013; Johannes et al. 2015). Interestingly, multipolarity can also occur without centrosome amplification. For example, defects in mitotic spindle maintenance can lead to an aberrant yet functional spindle pole (Logarinho et al. 2012).

### Chromosome Cohesion Defects

The cohesin protein complex maintains centromeric cohesion between sister chromatids until anaphase onset, and unscheduled loss of cohe-

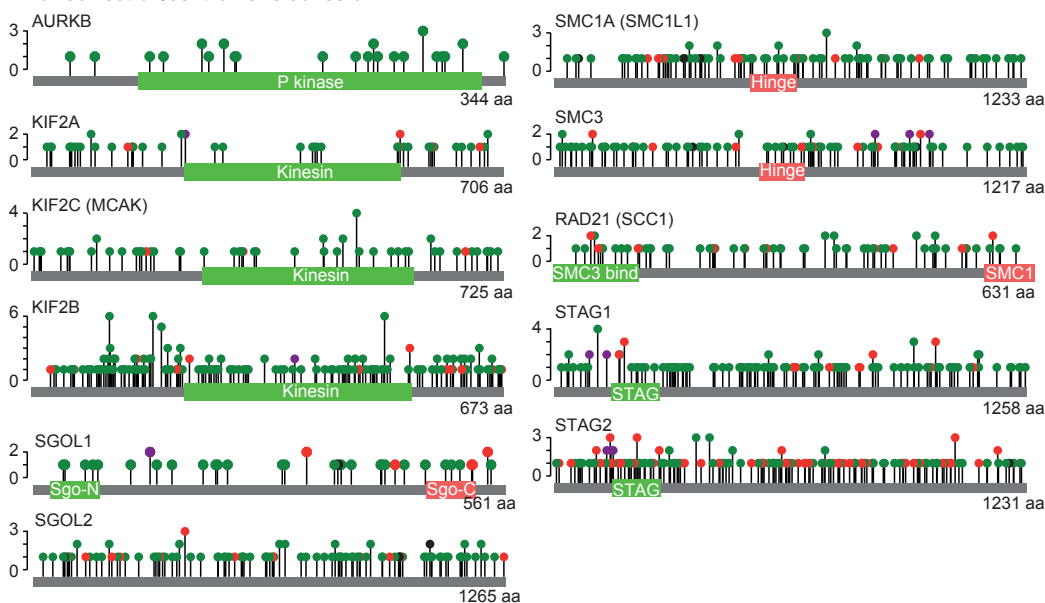
sion was shown to cause CIN (Jallepalli et al. 2001; Barber et al. 2008). Truncating mutations in the cohesin complex subunit STAG2 were identified in cancer cell lines and tumor samples and account for ~39% of STAG2 mutations reported in The Cancer Genome Atlas (TCGA) (Fig. 2) (Solomon et al. 2011). Introduction of a STAG2 inactivating mutations in a karyotypically stable cell line was sufficient to induce CIN, whereas correction of naturally occurring mutations reduced CIN in glioblastoma cell lines (Solomon et al. 2011). Because STAG2 is located on the X chromosome, complete gene inactivation can be achieved through a single mutation without the need for loss of heterozygosity (LOH). Although most but not all STAG2 truncating mutations lead to CIN and aneuploidy, the consequence of missense mutations causes various phenotypes, and the impact of cohesion complex mutations in cancer might also involve their general role of cohesion in transcription (Taylor et al. 2014; Kim et al. 2016).

Centromeric cohesion maintenance during mitosis also requires the action of Shugoshin 1 (SGO1) at the inner centromere (Watanabe 2005; Marston 2015). Recently, it was shown that SGO1 maintenance at the inner centromere was impaired in CIN cells when centromeres come under tension at metaphase. Interestingly, SGO1 recruitment at the centromere required both cohesin and the histone mark H3K9me3, and several CIN cell lines were found to be defective in one or both pathways (Tanno et al. 2015). Owing to its interaction with AURKB-containing chromosomal passenger complex, SGO1 mislocalization impairs error correction. This supports the notion that cohesion defects might cause CIN through stabilization of k-MT attachment errors (Kleyman et al. 2014; Tanno et al. 2015).

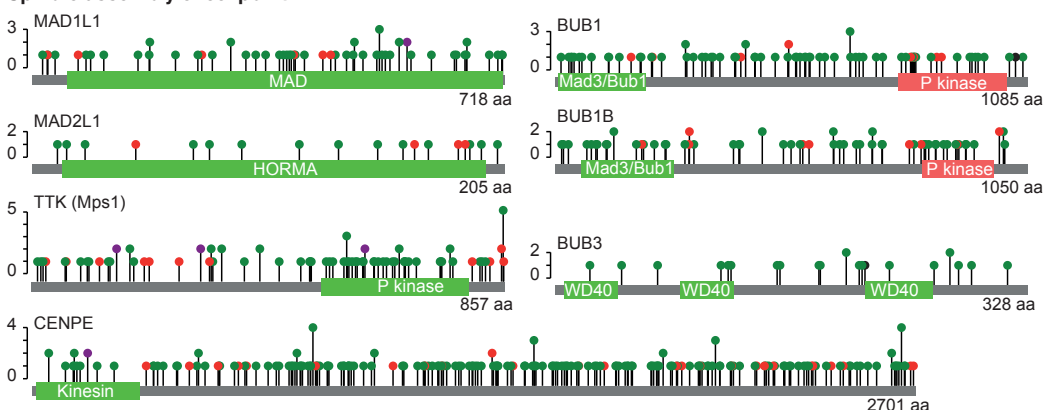
### CIN-Causing Mutations

The general view when considering the underlying genetic causes of wCIN is that direct mutations in any single gene responsible for segregation fidelity are rare. This conclusion is based on studies in which the mutational status of

### Error-correction/centromeric cohesion



### Spindle assembly checkpoint



**Figure 2.** Mutations identified in genes important for chromosome segregation fidelity. Lollipop plots depicting the location and nature of mutations identified in 6807 tumors (32 cancer types) sequenced by The Cancer Genome Atlas (TCGA). Pathway analyses might shed new light on the frequency of chromosomal instability (CIN)-causing defects and the underlying mutations driving CIN, which will require a deeper understanding of the consequence of missense mutations. Many mutations occur in functional domains: for example, 33/67 (49%) of missense mutations in TTK (Mps1) occur in the kinase domain. Green dot, missense mutation; red dot, truncating mutation (nonsense, splice site, and frameshifts); purple dot, various mutation types. (Data and diagrams modified from cBioportal [Cerami et al. 2012; Gao et al. 2013].)

CIN genes were analyzed independently in relatively small cohorts. This finding is perhaps expected for such a complex process in which mild disruptions in the activity of most proteins involved could elevate chromosome missegregation rates. Pan-cancer analyses based on

whole-genome or whole-exome sequencing data will enable the identification of low-frequency drivers, and allow pathway-driven rather than gene-centric analyses. This might reveal that tumors bear a mutation in a CIN-causing gene more frequent than previously anticipated

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(Leiserson et al. 2015). In Figure 2, we summarize the mutational data for some genes involved in error correction/cohesion and the SAC collected from 6807 tumors of diverse origins sequenced by TCGA (Cerami et al. 2012; Gao et al. 2013). As discussed further below (the section on Determinants of CIN Propagation), mildly disrupting mutations may be more potent CIN drivers than deleterious mutations, by causing low levels of CIN more likely to be viable and tolerated. A deeper understanding of the impact of missense mutations in candidate CIN drivers is also required, and genome-editing techniques will now enable us to explore the functional impact of these mutations in unprecedented detail. Epigenetic silencing may also emerge as a frequent CIN driver, as has been reported for BUB1B (Park et al. 2007; Haruta et al. 2008; Landau et al. 2014).

Mutations in common oncogenes and tumor suppressors are also associated with CIN. For example, deregulation of the RB-E2F and MYC pathways, or oncogenic Ras signaling has been reported to cause to aberrant expression of SAC genes, perturb microtubule dynamics, and cause centrosome amplification and cohesion defects (Hernando et al. 2004; Adon et al. 2010; Zeng et al. 2010; Kamata and Pritchard 2011; Schwartzman et al. 2011; Orr and Compton 2013; Manning et al. 2014; reviewed in Duijif and Benezra 2013; Orr and Compton 2013). Mutations in numerous tumor-suppressor genes have been shown to cause CIN, such as APC and WNT signaling pathway components (CHK1 and CHK2-BRCA1) (Zachos et al. 2007; Stolz et al. 2010; Orr and Compton 2013). The convergence of these various pathways on processes directly controlling segregation fidelity lead to the concept of oncogene-induced mitotic stress (Malumbres 2011; Duijif and Benezra 2013).

### INTERPLAY BETWEEN GENOMIC INSTABILITY AND CHROMOSOMAL INSTABILITY

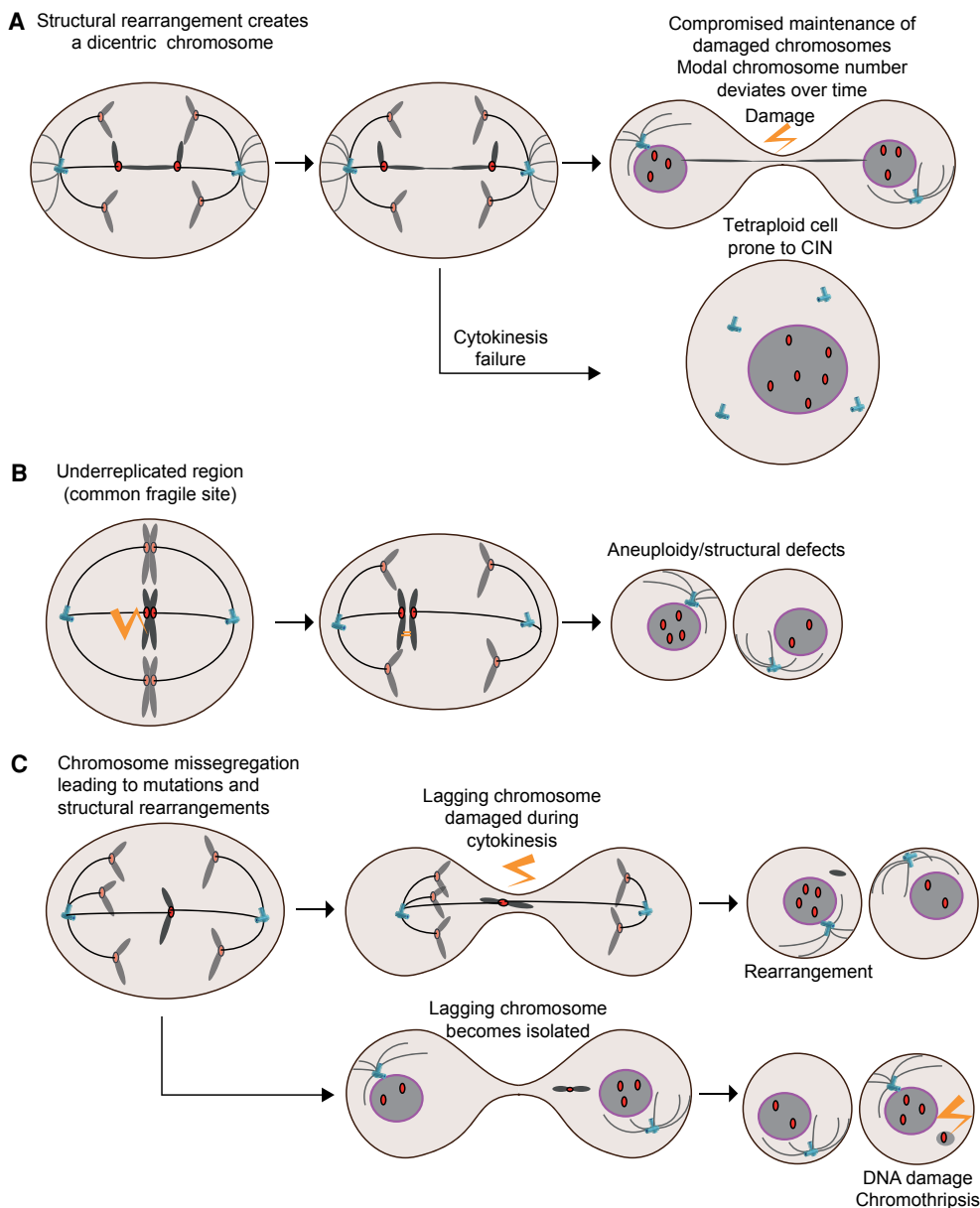
Cancer cells almost invariably display a combination of numerical (aneuploidy) and structural aberrations (translocations, deletions), as

well as defects affecting accurate replication or repair of damaged DNA (GIN). There is now accumulating evidence describing how the various types of instabilities can impact on each other, explaining at least in part how any of these triggers can pave the way for cancer cells to acquire such complex genetic compositions.

### Genomic Instability Fuels CIN

Premitotic defects arising from DNA replication fork collapse (replication stress) can result in double strand breaks (DSBs) especially at common fragile sites, which are late-replicating loci in the genome. Replication stress leads to rearrangement and chromosome breakage during the repair process especially when coupled to DNA repair defects (Halazonetis et al. 2008). As a result, replication-stress-induced defects can result in acentric chromosome fragments that will not be captured by the mitotic spindle or in dicentric chromosomes leading to chromosome bridges at anaphase (Fig. 3A) (Gissels-son 2008). These structural defects constitute a large fraction of mitotic aberrations during anaphase found in CIN colorectal cancer cells, and alleviation of replication stress using nucleoside complementation reduced their frequency indicative of their *de novo* formation during each phase of DNA replication (Burrell et al. 2013a). Transient induction of replication stress with aphidicolin also caused a deviation in chromosome number, a defining feature of CIN, when analyzed by clonal fluorescence in situ hybridization (FISH) using centromeric probes (Burrell et al. 2013a). Remarkably, it was recently shown that DNA replication occurs at common fragile sites during mitosis, presumably in a last attempt to complete DNA replication before cell division, and failure to do so causes chromosome missegregation as revealed by centromeric FISH (Minocherhomji et al. 2015). Replication stress can therefore lead to the missegregation of an entire, yet probably damaged, chromosome (Fig. 3B). Replication stress in precancerous lesions might therefore allow the onset of CIN in some tumors (Gorgoulis et al. 2005).

Apart from common fragile sites, telomeres are also intrinsically sensitive to replication



**Figure 3.** The interplay between whole-chromosome instability (wCIN), structural CIN (sCIN), and genomic instability (GIN). (A) Structural rearrangements caused by replication stress before mitosis created dicentric chromosomes. These can result in ultrafine bridges devoid of nuclear membrane and subject to extensive DNA damage and rearrangements akin to chromothripsis or hypermutation through kategais and severing of the chromosome by exonucleases. Dicentric chromosomes can also prevent cytokinesis, as observed following telomere dysfunction. (B) Replication stress at common fragile sites triggers DNA replication during mitosis. Failing to complete DNA replication can lead to nondysjunction and aneuploidy. (C) Whole-chromosome missegregation can result in the trapping of the lagging chromosome during cytokinesis, causing DNA damage and structural aberrations. Alternatively, lagging chromosomes can become isolated in a micronucleus, in which the lack of nuclear membrane renders it susceptible to extensive DNA damage and chromothripsis. Damaged micronuclei can reintegrate the other chromosomes in the following division and be propagated.

stress. Telomere maintenance is crucial for ongoing cell proliferation, and failure to do so results in telomere attrition and eventually activation of a DNA damage response, which halts cell proliferation (telomere crisis). Evidence of transient telomere crisis during early stages of breast cancer, followed by stabilization of shorter telomeres through telomerase reactivation, was shown for several cancers including prostate (Meeker et al. 2002), colon (Rudolph et al. 2001), and pancreas (Chin et al. 2004; Feldmann et al. 2007). Interestingly, telomere dysfunction can result in genome-doubling through cytokinesis failure or rereplication (endoreduplication) (Fig. 3A). The resulting tetraploid cells displayed hallmarks of CIN and were tumorigenic (Davoli et al. 2010; Davoli and de Lange 2012). Dysfunctional telomeres also cause chromosomal rearrangements, including dicentric chromosomes, which result in chromosome bridges during division (Fig. 3A). Dicentric chromosomes were shown to be prone to extensive localized mutagenesis (kataegis, bearing a mutation signature characteristic of the APOBEC family of cytidine deaminase enzymes) and chromothripsis (the random rearrangement of an entire chromosome or chromosome segment) (Stephens et al. 2011; Leibowitz et al. 2015; Maciejowski et al. 2015). Chromosome bridges were found to be coated with RPA proteins, indicative of single-stranded (ssDNA) exposure (the substrate for APOBEC enzymes), and the bridge actively processed by an exonuclease (Maciejowski et al. 2015). This is consistent with the detection of DNA damage markers on chromosome bridges that can form following SAC dysfunction (Janssen et al. 2011b). Segregation error defects, by physically isolating chromosomes or part thereof, can thus provide an explanation for the observation of mutational clusters, and for segmental structural rearrangements.

### Chromosome Missegregation and Genomic Instability

A large body of evidence suggests that whole-chromosome missegregation and aneuploidy can in turn drive further GIN. Studies in yeast

revealed aneuploid strains are prone to further segregation errors, display a mutator phenotype, and acquire chromosomal rearrangements (Pavelka et al. 2010; Sheltzer et al. 2011). A single chromosome missegregation event can thus trigger hallmarks of full-blown GIN. Similarly, engineered human cell lines bearing an extra chromosome acquire segmental aneuploidy and rearrangements, an event most likely driven by replication stress (Passerini et al. 2016). In diploid cells, lagging chromosomes caused by SAC inhibition or microtubule attachment defects can also get trapped and damaged during cytokinesis, resulting in translocations (Janssen et al. 2011b). Whole-chromosome missegregation (wCIN) and aneuploidy can therefore lead to structural chromosomal aberrations (Fig. 3C). Consequently, CIN allows cells to rapidly explore various compositions of whole-chromosome and segmental aneuploidies, and the combination of oncogenes and tumor-suppressor genes on each chromosomal fragment most likely drives karyotypic selection (Davoli et al. 2013).

Chromosome missegregation can result in the physical isolation of one or few chromosomes into micronuclei, in which they are prone to extensive DNA damage because of severe nuclear envelope defects (Fig. 3C) (Crasta et al. 2012; Hatch et al. 2013). DNA from micronuclei can be reincorporated into the main chromosome mass and transmitted during the following division (Crasta et al. 2012). This provides a mechanism whereby a single chromosome segregation error results in the acquisition of mutations without prior defects in genome maintenance, and can therefore be a precursor of GIN. By combining live-cell imaging and single-cell sequencing, Pellman and colleagues showed that chromothripsis can occur in micronuclei, providing a simple mechanism to massively rearrange entire chromosomes while leaving the others intact (Zhang et al. 2015). Interestingly, chromothripsis can also explain the formation of double-minute chromosomes, short circular acentric DNA fragments that are often amplified and contain oncogenes. Pan-cancer studies have detected evidence of chromothripsis in 2%–5% of samples, with higher



prevalence in some cancer types such as glioblastoma and bone cancers (Stephens et al. 2011; Zack et al. 2013; Rode et al. 2016). CIN thus creates at least two situations, namely, chromosome bridges (including ultra-fine bridges) (Liu et al. 2014) and micronuclei, in which loss of nuclear envelope integrity renders chromosomes prone to GIN (Hatch et al. 2013).

## DETERMINANTS OF CIN PROPAGATION

### Sustainable versus Lethal Levels of CIN

CIN is generally linked to poor outcomes in cancer (McGranahan et al. 2012). However, there is mounting evidence that the level of CIN determines its impact on tumor progression, in which a trade off at each division must be reached between the selective advantages that may be acquired versus the risk of acquiring an unviable karyotype. The concept of a “just right” level of CIN was crystallized by Lengauer and colleagues who proposed that CIN can create a breath of clones, which may by chance, possess the right characteristics to cross various selection barriers. However, this is only if CIN itself does not impair cell viability (Cahill et al. 1999). This idea is now supported by evidence from sequencing data and experimental models.

Genomic analysis of human data revealed a nonlinear relationship between the extent of chromosomal rearrangements and patient outcome. Patients whose tumor displayed either the lowest or highest instability (measured as the weighed Genome Instability Index, wGII) had better outcome than patients with intermediate levels of GIN in breast, ovarian, NSCLC, and gastric adenocarcinoma (Birkbak et al. 2011; Roylance et al. 2011). Extreme CIN was also linked to improved outcome in a multivariate analysis performed on a cohort of more than 1100 ER-negative breast cancer patients in which CIN was derived from centromeric FISH analysis (Jamal-Hanjani et al. 2015). More recently, similar conclusions were reached in a pan-cancer analysis exploiting exome data from 1165 tumors across 12 tumor types and further validated with SNP-array data sets from more than 2000 samples (Andor et al. 2016). By examining

the correlation between survival and the genome fraction affected by copy number alterations, the investigators found better outcome in patients when either <25% or >75% of the tumor genome was affected by copy-number variants. Tumor progression is thus more likely to occur with moderate levels of karyotypic variations.

Mouse models of CIN also support the hypothesis that only moderate CIN levels are viable and can promote cancer, whereas extreme CIN is unviable. Homozygous deletion of many CIN-causing genes is embryonically lethal in mice (reviewed in Simon et al. 2015), whereas heterozygous mice show moderate levels of aneuploidy with a higher incidence of cancer (either spontaneous, in a sensitized background or with carcinogens). In a landmark study using CENPE<sup>+/-</sup> mice, Cleveland and colleagues showed that CIN promotes tumorigenesis in tissues with low endogenous levels of aneuploidy (lung and lymphocytes), but is tumor suppressive in the liver, which displays high levels of endogenous segregation errors (Weaver et al. 2007). Exploiting the tumor suppressive function of excessive CIN is the basis for using SAC inhibitors to target cancer cells and could contribute to the cytotoxic effect of paclitaxel (Janssen et al. 2011a; Zasadil et al. 2014; Drost et al. 2015; Lee et al. 2016).

### Aneuploidy Tolerance Mechanisms

Aneuploidy has been well documented to be detrimental to overall organismal development and cell fitness in virtually every species studied, from yeast, plants, and mammals (Siegel and Amon 2012). Studies in yeast and mouse showed that the presence of an extra chromosome impairs cell proliferation regardless of the extra chromosome involved and was associated with altered metabolism (Torres et al. 2007; Williams et al. 2008). The presence of an extra chromosome and segmental aneuploidies results in a proportional perturbation in transcript and protein levels for most encoded genes (Rancati et al. 2008; Pavelka et al. 2010; Torres et al. 2010; Stinglele et al. 2012; Dephoure et al. 2014), although some gene dosage compensation has been reported. Increased pressure on

the translation machinery, protein quality control mechanisms, and stoichiometric imbalances brought about by aneuploidy, contribute to a global stress response believed to hamper cell proliferation (Santaguida and Amon 2015). Aneuploid cells displayed greater sensitivity to drugs that further impair these pathways, exposing vulnerabilities of aneuploid cells that could be exploited therapeutically (Tang et al. 2011).

TP53 status appears to be an important determinant of CIN propagation, and analysis of TCGA data revealed it is the only gene significantly associated with CIN (Burrell et al. 2013a). In Barrett's esophagus, TP53 LOH precedes tetraploidization and aneuploidy (Barrett et al. 1999; Lai et al. 2007). TP53 is stabilized following chromosome missegregation and results in a p21(CDKN1A)-dependent cell-cycle arrest (Thompson and Compton 2008, 2010). Interestingly, mice expressing a p53 mutant (R175P in human) defective for apoptotic induction but that retains cell-cycle arrest functions, showed delayed lymphomagenesis compared with TP53<sup>-/-</sup> mice and only gave rise to diploid tumors, an effect which was entirely dependent on p21 induction (Liu et al. 2004; Barboza et al. 2006). The mechanistic basis linking chromosome segregation errors and p53 stabilization is still elusive, but it is likely that several upstream signaling pathways are necessary, one of which being the p38 MAPK pathway (Thompson and Compton 2010; Kumari et al. 2014). p53-independent pathways might also act in parallel. Mutational timing analysis in a pan-cancer dataset revealed that TP53 mutations are predominantly clonal and as such tumors might be primed to be CIN tolerant early in the etiology of cancer (McGranahan et al. 2015). TP53 mutations are also significantly associated with genome doubled tumors and occur before genome duplication in >90% of tumors (McGranahan et al. 2015). 17p deletions encompassing the TP53 locus affect ~70% of human cancers and result in co-loss of an AURKB allele, potentially facilitating CIN and tolerance in a single event (Fernandez-Miranda et al. 2011; Liu et al. 2016). The importance of TP53 in blocking proliferation of tetraploid

cells, by imposing a G1 arrest, has been well documented (Andreassen et al. 2001; Fujiwara et al. 2005; Davoli and de Lange 2012; Ganem et al. 2014). TP53 disruption is therefore thought to render cells permissive for CIN by relieving the stress associated with acute segregation errors and by allowing the expansion of genome doubled cells, which are intrinsically more prone to CIN.

Genome-doubling represents arguably one of the most underestimated precursor of CIN in human cancers. Newly formed tetraploidization cells display CIN leading to extensive karyotypic heterogeneity and chromosomal rearrangements in various systems, and can initiate tumor formation in mice (Fujiwara et al. 2005; Davoli and de Lange 2012; Dewhurst et al. 2014). Tetraploidization and CIN are also early events in esophageal cancer, such as in Barrett's esophagus (Barrett et al. 1999; Lai et al. 2007; Davoli and de Lange 2011; Murugaesu et al. 2015). Recent large-scale copy number analyses of human cancers revealed that an ancestral whole-genome doubling event is detected in ~37% of tumors (11% to 64% depending on cancer type) (Cibulskis et al. 2012; Zack et al. 2013; Dewhurst et al. 2014). Such tumors display a much higher rate of copy-number gain and losses, most of which is acquired post genome-doubling (Zack et al. 2013; Dewhurst et al. 2014). Experiments with various cell lines showed that tetraploid cells are also much more tolerant to segregation errors than their diploid precursors, without prior mutations (Dewhurst et al. 2014; Kuznetsova et al. 2015). Deleterious genetic alterations are most likely buffered in a tetraploid background, providing a genetic environment into which a wider range of genetic makeups can be explored.

### Sporadic CIN Episodes and Compensation

CIN is unlikely to follow a linear trajectory in which the level of instability may fluctuate dramatically during tumor development. As a result, CIN may itself be dynamic during tumor evolution and subject to various compensatory mechanisms, an area that remains poorly explored. Although genomic imbalances globally

translate into proportional differences on transcript and protein abundance, as much as 20% of the genes on an extra chromosome may not show aberrant expression (Torres et al. 2007; Williams et al. 2008; Pavelka et al. 2010; Stinglee et al. 2012). This regulation might take place on a gene-specific basis in aneuploid cells, and could involve translational compensation (Dassi et al. 2015). Cells could also compensate for the sudden loss of CIN inducing genes, especially for those whose loss would lead to excessive CIN, by developing alternative coping mechanisms. This phenomenon has been described following Securin (PTTG1) homozygous deletion, which causes extensive CIN, followed by adaptation to the lack of securin and genome stabilization (Pfleghaar et al. 2005). SAC activity is also dispensable after the combined loss of at least two E2 ubiquitin ligases involved in mitotic progression (Wild et al. 2016). Cells can thus respond to intensive CIN episodes through buffering mechanisms, to promote survival.

## EXPLORING THE FITNESS LANDSCAPE THROUGH CIN

### Aneuploidy Drives Adaptation

Elegant experiments in yeast have provided evidence that aneuploidy can drive phenotypic adaptation. They revealed that although aneuploidy hampers cell proliferation under homeostatic conditions, it gives rise to clones with growth-promoting properties when exposed to chemotherapeutic and antifungal drugs, and drives the acquisition of entirely new traits on deletion of a gene thought to be essential for cell division (Rancati et al. 2008; Pavelka et al. 2010). Aneuploidies involving random combinations of chromosomes offer further possibilities to modulate independent pathways through additive and synergistic effects, producing complex phenotypes (Potapova et al. 2013). A striking observation is that aneuploid cells invariably show a greater phenotypic variance in response to cytotoxic agents, even when aneuploid cells were overall more sensitive to a drug than euploid cells, rare aneuploid clones

showed greater viability (Chen et al. 2015). Similarly, increased polyploidy greatly accelerates adaptation in budding yeast grown in raffinose as sole carbon source (Selmecki et al. 2015). Tetraploid cells not only acquired significantly more mutations than diploid or haploid cells, but only tetraploid clones almost inevitably displayed whole-chromosome and segmental aneuploidies (Selmecki et al. 2015). Remarkably, copy gain of chromosome XIII accelerated adaptation in tetraploid clones but was detrimental when occurring in diploid cells. Tetraploidy thus provides a more permissive background in which a greater breath of karyotypic and mutational variations can be explored, increasing the probability of acquiring growth-promoting properties.

### CIN in Tumor Progression

CIN remains underestimated when analyzing mechanisms of tumor progression and relapse. Indeed, copy-number analyses on single biopsies poorly capture the extent of cell-to-cell variations and rather reflect pervasive copy number states. High degree of intratumor heterogeneity results in neutral copy number being observed for most chromosomes. In addition, analyses of highly aneuploid tumors (Sotillo et al. 2010) or mixtures of distinct aneuploid populations (Chen et al. 2015) create an illusion of diploidy. Multiregion sampling provides a better resolution and revealed in several studies that CIN onset preceded subclonal expansion and metastatic dispersion in several cancers, because chromosomal aberrations were shared between all regions, or occurred after genome-doubling (Campbell et al. 2010; de Bruin et al. 2014; Kim et al. 2015). Single-cell analysis of two paired primary and metastatic breast carcinomas revealed ongoing CIN in the primary tumor with the predominance of a highly aneuploidy clone, which seeded the metastasis (Navin et al. 2011). Multiregion sequencing also revealed that most metastatic copy number and structural variants arose from a primary tumor subclone, but with evidence of persistent CIN during the metastatic process (Yates et al. 2015).

### CIN Drives Recurrence and Metastasis

Seminal experiments using an inducible K-Ras-driven model of lung cancer showed that concomitant inducible CIN induction (through MAD2 overexpression) during tumor development greatly increases recurrence following K-Ras oncogene withdrawal (Sotillo et al. 2010). In this system, K-Ras/MAD2 tumors displayed CIN and enhanced the aggressiveness, but tumor regression was just as profound following oncogene withdrawal. However, 11/24 of tumors from K-Ras/MAD2 mice recurred, as opposed to 0/25 for K-Ras tumors after oncogene withdrawal. As pointed out by the investigators, this situation is reminiscent of Bcr-Abl-driven CML tumors, which invariably respond to imatinib; however, recurrence is significantly higher in tumors initially showing CIN (Cortes and O'Dwyer 2004). The fact that MAD2 transgene expression is dispensable for tumor maintenance reflects an important feature of CIN in general. Once initiated, CIN provides a pool of highly diversified cells, which are prone to acquire further numerical and structural aberrations. This relieves the selective pressure to maintain the initial genetic lesion, and is therefore not subject to oncogene addiction. Mouse models of skin and pancreatic cancers have similarly shown that CIN drives metastatic progression (Hingorani et al. 2005; McCreery et al. 2015). In human tumors, increase in CIN has been linked to metastatic progression in prostate cancer, pancreatic cancer, breast cancer, colorectal cancer, and renal cell carcinoma (reviewed in Turajlic and Swanton 2016).

### CIN Contributes to Drug Resistance

CIN has long been associated with multidrug resistance (Duesberg et al. 2007; Lee et al. 2011; McGranahan et al. 2012). Tetraploid cells, by virtue of their higher CIN rate and tolerance (Dewhurst et al. 2014), also display greater resistance to a broad range of compounds (Lee et al. 2011; Dewhurst et al. 2014; Kuznetsova et al. 2015). One explanation for this phenomenon is that CIN allows the maintenance of various karyotypes, and the one(s) conferring a proliferative advantage expand on selection.

For example, in a longitudinal study of CLL, increased genomic complexity on recurrence was observed more frequently amongst patients who received therapy (Ouillette et al. 2013). In medulloblastoma, profound divergence was observed posttherapy, yet the clone representing <5% of the primary tumor driving recurrence featured aneuploidy and GIN (Morrissy et al. 2016). Detailed phylogenetic reconstruction of chemotherapy resistant breast cancer also uncovered the presence of the resistant subclones before treatment, showing that chemotherapy did not drive the acquisition of mutations (Yates et al. 2015). Instead, resistance could be traced back to the expansion of a preexisting clone with persistent GIN.

### CONCLUDING REMARKS

Although wCIN is a pervasive feature of human tumors, it is challenging to identify the genetic and/or epigenetic cause(s) of wCIN in individual tumors. This might be predictable given that CIN can be brought about by a multitude of defects and its propagation does not require maintenance of the CIN initiating lesion. An alternative explanation is that mutations that cause a high level of CIN might be excessively deleterious, even in a genetic background permissive to aneuploidy, such that CIN might be driven mainly by partially deleterious mutations. For similar reasons, known oncogenes might be potent CIN drivers because they moderately affect the expression and function of genes involved in genome maintenance. Nevertheless, CIN enables the exploration of a phenotypic landscape in a way that cannot be achieved with an increased mutation rate alone (Mroz et al. 2015). A deep understanding of the underlying causes of CIN and the context enabling its propagation is still lacking and needs thorough longitudinal studies to fully understand its importance. Stochastic chromosome missegregation in normal cells, although rare (one error per 100–500 divisions), explains the presence of aneuploid cells in various healthy tissues (Knouse et al. 2014) and could represent early precursors of CIN. Understanding genome dynamics during therapy response

and resistance might also offer new strategies to ambush cancer cells by forcing them through an “evolutionary trap” that could make them exquisitely sensitive to a second treatment (Chen et al. 2015). The development of sensitive approaches for copy-number analysis should now prompt study design aimed at tracking more precisely the onset of CIN in tumors and targeting of CIN for therapeutic benefit.

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